Chronic, Active Inflammation in Patients With Failed Total Knee Replacements Undergoing Revision Surgery

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ABSTRACT: Chronic pain and restricted knee motion is a significant problem following the total knee arthroplasty (TKA). The molecular pathogenesis of pain post-TKA is not known and no targeted therapeutic intervention is available. The aim of this study was to investigate whether pro-inflammatory mediators are elevated in revision knee patients, indicating an active, ongoing inflammatory process that may contribute to pain. Twelve key markers (pro-inflammatory cytokines granulocyte-macrophage colony-stimulating factor (GM-CSF), interleukin 5 [IL-5], IL-8 and IL-10, chemokines CCL2, CCL3, CCL4, and CCL13, mediators of angiogenesis Flt-1, vascular endothelial growth factor, and cell migration vascular cell adhesion molecule 1 and intercellular adhesion molecule 1) were measured in knee tissue and synovial fluid (SF) from primary TKA (n = 29) and revision patients (n = 32). Indications for surgery were osteoarthritis (OA) for primary TKA, and component loosening (n = 11), stiffness (n = 11), laxity pattern (n = 8), or progression of OA in patella resurfacing (n = 3) for revision surgery. Pain levels (WOMAC score) were higher in revision than primary patients (p < 0.05). Time from primary to revision ranged from 8 months to 30 years (median 10 years). All markers were elevated in revision TKA; there was no trend toward decreasing levels with greater time from primary surgery for any marker studied in SF. Similar results were seen in knee tissue.

Keywords: fibrosis; inflammation; knee; arthroplasty; pain

Primary total knee arthroplasty (TKA) is one of the most commonly undertaken elective orthopedic procedures, with >98,000 performed in 2016 in the United Kingdom.1 While the majority of patients have a successful outcome, with reduced pain and improved function,2 a significant proportion (10–20%) develop problems including joint stiffness, reduced range of knee motion and pain, which can ultimately require revision surgery.3 Extensive work has been undertaken to identify patient factors (e.g., male/female sex, co-morbidities, mental health, and preoperative patient expectations)4 and surgical factors (e.g., implant design and surgical technique)5 that predict poor outcome following TKA. Increasingly, however, the role of patient biology as a potential driver of pain, stiffness and dissatisfaction is being considered.6 A relationship between levels of pro-inflammatory mediators (interleukin 6 [IL-6], IL-8, CCL2, C-reactive protein [CRP]) and pain levels in osteoarthritis has been well described.7,8 However, our understanding of the role of these mediators in pain and stiffness following TKA is more limited. Gandhi et al.9 studied 28 patients undergoing TKA and found that higher preoperative levels of tumor necrosis factor α (TNF-α), matrix metallopeptidase 13, and IL-6 in synovial fluid (SF) were associated with reduced improvement in pain scores 2 years following TKA. Conversely, Zietek et al.10 measured TNF-α levels in SF taken at time of primary surgery, and showed that higher levels of TNF-α were positively correlated with improved pain scores at 6 weeks post-surgery. Ugraş et al.11 measured SF levels of IL-6 and CRP 24 h postoperatively (sampling through a drainage tube placed intraoperatively) and found that higher levels of IL-6 were associated with slower recovery. These studies demonstrate uncertainty currently surrounding the role of pro-inflammatory cytokines in pain generation following TKA.

We and others have previously shown that the post-TKA joint is associated with the persistence of myofibroblasts and extensive fibrotic remodeling.12,13 More recently, we have investigated the expression of 39 soluble inflammatory proteins in SF, fat pad (FP), and synovial membrane (SM) from patients undergoing either primary or revision surgery.14 There was elevated expression of a significant number of these markers in each anatomical location: n = 26/39 markers in SF, n = 22/39 in infrapatellar FP and n = 10/39 in SM in revision versus primary knees.
The markers investigated in this study fulfilled two inclusion criteria; (i) they had previously been found significantly elevated in SF in revision TKR patients; and (ii) they are well characterized as mediators of fibrosis, inflammation, and/or pain. Specifically we were interested in pro-inflammatory and immune cell recruitment mediators, including the CCL family of chemokines. GM-CSF is produced by fibroblasts, promotes neutrophil and macrophage proliferation and maturation, and is upregulated immediately postoperatively in serum following TKA. IL-5 is a B-cell activating cytokine elevated in serum immediately following total knee replacement surgery that drives fibrosis via Th2 B cell development and myofibroblast activation. IL-8, a CXC chemokine involved in recruitment of neutrophils, has a positive correlation with IL-1 expression, suggesting that fibroblasts exposed to IL-1 in the surgical knee may be a source of IL-8. IL-10 has been demonstrated to drive lung fibrosis when overexpressed, and this process is dependent on CCL2 signaling and is elevated in serum following knee surgery. CCL2 levels correlate positively with patient’s pain levels following primary TKA, suggesting a role for CCL2 in pain pathogenesis, and recruitment of monocytes by knee fibroblasts in vitro is dependent on CCL2 signaling. CCL3 and CCL4 are elevated immediately following knee surgery and CCL13 is a key mediator of immune cell recruitment in chronic inflammation. Intercellular adhesion molecule 1 (ICAM-1) is an adhesion molecule expressed by macrophages and other leukocytes and vascular cell adhesion molecule 1 (VCAM-1) is expressed by activated endothelium and facilitates trans-migration of immune cells into tissues. Soluble vascular endothelial growth factor (VEGF) and Flt-1 are regulators of vessel growth; VEGF directly stimulates new vessel formation and growth of existing vessels, signaling through Flt-1.

However it is not known whether the inflammatory response post-TKA is acute and resolving, or whether this process is chronic. To test this we examined the levels of key pro-inflammatory mediators in SF and tissue of patients undergoing revision surgery and compared this to time elapsed from their primary TKA procedure.

Results presented here suggest that key, active pro-inflammatory and matrix deposition markers are elevated in patients undergoing revision surgery, demonstrating the presence of active and ongoing chronic inflammation. These findings provide rationale for targeted biological therapy in knee pain and stiffness, even several years after the primary surgery. Furthermore, these key mediators could be molecular targets in patients with unexplained knee pain following TKR.

METHODS
Patient Recruitment and Ethics
This research has been conducted following ethical approval through the Newcastle Biobank (17/NE/0361).

Tissue Collection and Patient Stratification
All patients undergoing revision surgery for failed primary TKA were included in the study over a two-year period with infection as the only exclusion criterion. Level of evidence—III, retrospective cohort study comparing disease and control populations. SF, infrapatellar fat pad (FP), and synovial membrane (SM, suprapatellar pouch) were collected from patients undergoing either primary (n = 29) or revision (n = 33) TKA at the Freeman Hospital, Newcastle upon Tyne (Table 1). Patient reported outcome measures (Western Ontario & McMaster Universities Osteoarthritis Index, WOMAC scores) were recorded as routine practice in the Freeman hospital joint registry at the time of surgery. Revision TKA patients were stratified as those with osteolysis and loose components (n = 11), clinical diagnosis of fibrosis with loss of movement (n = 11), primary laxity pattern with functional instability (n = 8, see Supplementary Methods for details), and progression of osteoarthritis (previous patellofemoral resurfacing) (n = 3). The clinical definition of joint fibrosis and method for exclusion of infection was as previously described (Supplementary Methods). Types of knee replacement in the revision cohort and surgical technique are described in Supplementary Methods. Histological analysis was performed on 9/33 patients (see Supplementary Methods).

Infrapatellar Fat Pad and Synovial Membrane
Tissue was stored at 4°C in tissue culture media containing 10% fetal calf serum, 1% L-glutamine, 100 units/ml penicillin, and 100 μg/ml streptomycin before processing. Tissue was homogenized in radioimmunoprecipitation assay buffer supplemented with protease and phosphatase inhibitors using a bead homogenizer (TissueLyserII; Qiagen, Germantown, MD). Homogenized samples were normalized to 1 mg/ml total protein (protein concentration measured using a BCA protein assay (Pierce) as per manufacturer’s instructions) and stored at −80°C before analysis.

Multi-Array Protein Assays
Not all patients had fluid and both tissue samples collected at the time of surgery. SF (primary n = 21 patients, revision n = 24 patients), and tissue homogenates from infrapatellar FP (primary n = 28 patients, revision n = 32 patients) and SM (primary n = 29 patients, revision n = 32 patients) were assessed for expression of 12 protein markers using human V-Plex electrochemiluminescence detection kits from Meso-Scale Discovery as per manufacturer’s
Age regarded as significant.

Statistical Analysis
Differences in patient demographics were examined using Mann–Whitney U (Graph Pad, Prism, San Diego, CA, version 7). Time from primary TKA to revision was modeled using marker levels, age, and body mass index (BMI) using multiple linear regression (performed in R, available at www.r-project.org). Differences in marker levels between primary and revision patients, and between different revision groups were examined using Mann–Whitney U. Data are presented as mean ± standard error of the mean (SEM). 

RESULTS

Table 1. Patient Demographics

<table>
<thead>
<tr>
<th>Component</th>
<th>Primary (n = 29)</th>
<th>All Revisions (n = 33)</th>
<th>p Value</th>
<th>Osteolysis, Loose Components (n = 8)</th>
<th>Primary Laxity Pattern (n = 3)</th>
<th>Progression of OA/PFJ Replacement, Fibrosis (n = 11)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years, mean, SEM)</td>
<td>66 ± 2</td>
<td>68 ± 2</td>
<td>0.76</td>
<td>70 ± 3.2</td>
<td>67 ± 5</td>
<td>69 ± 6</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
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<tr>
<td>M:F 12:16</td>
<td>16 M:17 F</td>
<td></td>
<td>0.88</td>
<td>5 M:6 F</td>
<td>4 M:4 F</td>
<td>3 F</td>
</tr>
<tr>
<td>BMI (Mean, SEM)</td>
<td>33 ± 1</td>
<td>33 ± 1</td>
<td>0.97</td>
<td>34 ± 2</td>
<td>33 ± 2</td>
<td>30 ± 2</td>
</tr>
<tr>
<td>Time from primary to revision surgery (years, mean, SEM)</td>
<td>–</td>
<td>10 ± 1</td>
<td></td>
<td>16 ± 3</td>
<td>6 ± 1</td>
<td>6 ± 3</td>
</tr>
<tr>
<td>WOMAC pain scores (Mean, SEM)</td>
<td>30 ± 3</td>
<td>52 ± 7</td>
<td>0.001</td>
<td>65 ± 5</td>
<td>49 ± 4</td>
<td>34 ± 7</td>
</tr>
</tbody>
</table>

No difference was found comparing age, gender, or body mass index (BMI) in revision versus primary patients, or comparing groups according to indication for revision (p > 0.05, U test). Pain was significantly higher in revision versus primary patients (p < 0.001).

Statistical analyses were performed using the MSD Discovery Workbench analysis software.

No difference was found comparing age, gender, or body mass index (BMI) in revision versus primary patients, or comparing groups according to indication for revision (p > 0.05, U test). Pain was significantly higher in revision versus primary patients (p < 0.001). SEM, standard error of the mean.

No difference was found comparing age, gender, or body mass index (BMI) in revision versus primary patients, or comparing groups according to indication for revision (p > 0.05, U test). Pain was significantly higher in revision versus primary patients (p < 0.001). SEM, standard error of the mean.
Table 2. Synovial Fluid

<table>
<thead>
<tr>
<th>Marker</th>
<th>Primary (n = 21)</th>
<th>Mean ± SEM (pg/ml)</th>
<th>All Revisions (n = 24)</th>
<th>Mean ± SEM (pg/ml)</th>
<th>p Value</th>
<th>Significance</th>
<th>Primary vs. Revision TKA</th>
<th>Osteolysis, Loose Components (n = 11)</th>
<th>Mean ± SEM (pg/ml)</th>
<th>Primary Laxity Pattern (n = 7)</th>
<th>Mean ± SEM (pg/ml)</th>
<th>Progression of OA (PFJ Replacement, n = 1)</th>
<th>Mean ± SEM (pg/ml)</th>
<th>Clinical Diagnosis of Fibrosis (n = 7)</th>
<th>Mean ± SEM (pg/ml)</th>
<th>p Value Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>GM-CSF</td>
<td>0.5 ± 0.2</td>
<td>1.3 ± 0.3</td>
<td>&lt;0.01</td>
<td>**</td>
<td>1.8 ± 0.5</td>
<td>0.5 ± 0.2</td>
<td>0</td>
<td>1.6 ± 0.5</td>
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<tr>
<td>IL-5</td>
<td>0.4 ± 0.1</td>
<td>2.2 ± 0.8</td>
<td>&lt;0.001</td>
<td>****</td>
<td>1.6 ± 0.4</td>
<td>4.1 ± 2.7</td>
<td>0</td>
<td>1.2 ± 0.4</td>
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<tr>
<td>IL-8</td>
<td>80 ± 31</td>
<td>3,147 ± 1,552</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>6,296 ± 3,996</td>
<td>777 ± 215</td>
<td>61</td>
<td>1,907 ± 829</td>
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<tr>
<td>IL-10</td>
<td>0.8 ± 0.5</td>
<td>1.6 ± 0.4</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>2.6 ± 0.9</td>
<td>0.8 ± 0.2</td>
<td>0</td>
<td>1.2 ± 0.6</td>
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<tr>
<td>CCL2</td>
<td>417 ± 38</td>
<td>5,285 ± 1,004</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>5,902 ± 1,233</td>
<td>3233 ± 791</td>
<td>318</td>
<td>7,254 ± 2,824</td>
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<tr>
<td>CCL3</td>
<td>15.6 ± 3.3</td>
<td>177 ± 51</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>285 ± 125</td>
<td>96 ± 16</td>
<td>16</td>
<td>142 ± 54</td>
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<tr>
<td>CCL4</td>
<td>69.6 ± 8</td>
<td>258 ± 54</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>368 ± 131</td>
<td>213 ± 39</td>
<td>36</td>
<td>192 ± 51</td>
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<tr>
<td>CCL13</td>
<td>42.5 ± 20</td>
<td>220 ± 76</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>211 ± 92</td>
<td>67 ± 22</td>
<td>14</td>
<td>211 ± 92</td>
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<tr>
<td>Flt-1</td>
<td>97.6 ± 12</td>
<td>1575 ± 612</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>3,025 ± 1,512</td>
<td>391 ± 170</td>
<td>33</td>
<td>1,116 ± 443</td>
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<tr>
<td>VEGF</td>
<td>396.1 ± 46</td>
<td>1724 ± 268</td>
<td>&lt;0.0001</td>
<td>****</td>
<td>1,589 ± 220</td>
<td>1,081 ± 292</td>
<td>133</td>
<td>2,768 ± 680</td>
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<tr>
<td>VCAM-1</td>
<td>105,613 ± 7,290</td>
<td>134,770 ± 7,434</td>
<td>&lt;0.05</td>
<td>*</td>
<td>138,249 ± 10,066</td>
<td>151,817 ± 15,596</td>
<td>96,090</td>
<td>118,775 ± 13,418</td>
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<tr>
<td>ICAM-1</td>
<td>862,890 ± 12,439</td>
<td>150,915 ± 24,118</td>
<td>&lt;0.05</td>
<td>*</td>
<td>189,061 ± 39,854</td>
<td>168,010 ± 58,990</td>
<td>31,383</td>
<td>101,851 ± 18,441</td>
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</table>

Levels of markers in synovial fluid. Comparing revision vs. primary synovial fluid all markers were significantly upregulated (p ≤ 0.05, U test). No significant differences were seen comparing different indications for revision surgery (p ≥ 0.05).

GM-CSF, granulocyte-macrophage colony-stimulating factor; ICAM-1, intercellular adhesion molecule 1; IL, interleukin; VCAM-1, vascular cell adhesion molecule 1; VEGF, vascular endothelial growth factor.

*p < 0.05.
**p < 0.01.
***p < 0.001.
****p < 0.0001.
(ICAM-1, VCAM-1) and proangiogenic mediators (VEGF and Flt-1) also remain elevated. ICAM-1 was upregulated twofold in SF and FP \((p \leq 0.05, \text{Table 2})\). Both VEGF and Flt-1 are upregulated in revision SF \((\text{fourfold and 16-fold})\). Again, there was no reduction in level of these four markers with greater time from surgery in both fluid and tissue \((\text{Fig. 3})\).

No marker studied demonstrated a reduction in level with greater time from surgery \((\text{Figs. 1–3, tested in a linear regression model})\). This finding suggests a chronic type-2 inflammatory response in revision patients.

No Significant Differences in Inflammatory Markers Between Different Revision Groups
As we had a heterogeneous group of revision patients who were undergoing surgery for one of four different indications we examined possible differences in levels of inflammatory markers between different revision groups. We found no significant differences in any of the markers studied in SF, FP, or SM tissue \((p \geq 0.05 \text{ for all tests; example graphs are shown in Figure 4, data is presented in Tables 2, S3 and S4 [Supplementary Results]})\).

DISCUSSION
Knee pain and stiffness is a devastating complication following total knee replacement yet our understanding of the biology of the painful TKA remains incomplete. \(^6\) Previous studies have tested the hypothesis that the level of inflammation pre- or peri-operatively is correlated with clinical outcome and/or pain. \(^10,11\) Here, we have studied a group of patients undergoing revision surgery for all causes (having excluded infection). We report that in patients undergoing revision surgery, who have higher pain scores than primary TKA patients, there is elevation of key inflammatory mediators in SF and knee tissue. This is present irrespective of time from primary surgery, with low \(r^2\) and \(p\) values in multiple linear regression for all markers tested against time. These findings suggest that targeted modulation of chronic inflammation may offer therapeutic benefit in the treatment of pain following TKA.

Stiffness and joint fibrosis is a significant problem following TKA. We have previously shown that fibrotic, dense collagenous scar tissue is found in all patients undergoing aseptic revision surgery for failed primary TKA. \(^12\) The fibrotic tissue is characterized by the deposition of a dense, disorganized extracellular matrix of collagen \(^23\) populated by myofibroblasts. \(^12,13\) Although all revision knees in this cohort have joint fibrosis, presumably there are differences in the anatomic location and amount of fibrotic tissue present.

**Pro-inflammatory markers**

**Figure 1.** Elevated levels of pro-inflammatory mediators granulocyte-macrophage colony-stimulating factor (GM-CSF), interleukin 5 (IL-5), and IL-8 are maintained over time following primary total knee arthroplasty (TKA) in synovial fluid and tissue in patients with joint fibrosis. No significant change in the levels of pro-inflammatory mediators GM-CSF, IL-5, and IL-8 was found over time from primary TKA surgery in synovial fluid, fat pad, or synovial membrane tissue. The red line indicates the mean level in primary tissue for comparison. The black line represents the mean and black long dashed lines the 95% confidence interval. A linear regression model was used; \(r^2\) and \(p\) values are presented for each analysis \([\text{Color figure can be viewed at wileyonlinelibrary.com}]\).
We have not been able to determine whether fibrosis is a result of an ongoing inflammatory state caused by a failing primary TKA, or is normal scar tissue found post-TKA. Consequently differences in the molecular biology of the stiff TKA must be further investigated and dissected out from those seen in TKAs that have failed for other reasons (e.g., loosening, component malposition). Animal models of joint fibrosis have been developed and may provide further mechanistic insights into the disease pathogenesis. Histological analysis in this study showed polyethylene wear particles that could have a pathogenic role in driving chronic inflammation.

Treatment of joint fibrosis is currently limited to physical therapy. Patients resistant to nonoperative treatment require arthroscopic or more invasive surgical procedures to excise and remove the soft tissue contractures. Administration of three doses of intravenous hydrocortisone decreased systemic inflammation (measured by serum IL-6 levels) 24 h postoperatively. Patients receiving steroid had improved ROM at time of discharge, and although no long-term follow-up data is available, dampening inflammation may provide a potential clinical benefit.

Previous studies have measured pre- or peri-operative levels of TNF-α and IL-6. We found TNF-α threefold higher in revision versus primary SF, supporting a possible role in promoting pain and inflammation. However, IL-6 was not upregulated in revision knees suggesting that IL-6 is most useful as an early pre-/peri-operative biomarker.

In this cohort pain scores were higher in failed TKA patients than patients undergoing primary surgery for osteoarthritis. This demonstrates the challenges faced to achieve a satisfactory outcome following revision.

Figure 2. Elevated levels of Immune cell recruitment mediators CCL2, CCL3, and CCL4 in synovial fluid and tissue following primary total knee arthroplasty (TKA) in patients with joint fibrosis. No significant change in the levels of CCL2, CCL3, and CCL4 was found over time from primary TKA surgery in synovial fluid, fat pad, or synovial membrane tissue. The red line indicates the mean level in primary tissue for comparison. The black line represents the mean and black long dashed lines the 95% confidence interval. A linear regression model was used; $r^2$ and $p$ values are presented for each analysis [Color figure can be viewed at wileyonlinelibrary.com]
and is consistent with previously reported higher levels of chronic pain after revision TKA compared with following primary TKA.\textsuperscript{31}

Several limitations to the present study should be noted. Revision patients studied here were a heterogeneous group undergoing surgery for all causes. We have not been able to robustly identify differences in the molecular profile comparing different indications for revision. There is no previously published data on differences in inflammatory microenvironment of the post-TKA knee to allow a power calculation to be performed. Revision surgery is relatively infrequently required following TKA, and despite an unbiased patient selection strategy for our revision cohort to maximize the number of patients recruited to the study, with the only rule-out clinical condition being infection, we are likely to be underpowered to detect differences between different indications for revision surgery in this exploratory experimental study. We have compared revision patients to primary patients with advanced osteoarthritis; the ideal control group would be TKA patients with a well-functioning prosthesis and this will be tackled in future studies. Biomarker changes may be age-related, and although we controlled for age in multiple linear regression analysis we cannot exclude an effect of age in revision patients, particularly elderly patients who underwent primary surgery >20 years ago. We were unable to control for different analgesic regimens between patients, which could influence outcomes and were not able to differentiate neuropathic pain, which may have a different pathogenesis and is known to occur in a sub-set of TKA patients.\textsuperscript{32}

Data presented in this report suggest that the levels of key biologically active pro-inflammatory mediators are locally elevated at time of revision surgery many years after the initial procedure in failed TKAs. The painful,\textsuperscript{14} fibrotic\textsuperscript{12} post-TKA joint acquires a state of unresolved inflammation with a signature resembling that of a chronic allergic reaction.\textsuperscript{33} Markers with this temporal regulation include those involved in matrix formation (VEGF, Flt-1) and the inflammatory cytokines GM-CSF, IL-5 and IL-8, and cell recruitment chemokines CCL2, CCL3 and CCL4. Similar results were obtained in tissue and SF. This finding has implications when considering therapeutic intervention. The elevation of these markers indicates an active disease process, thus offering the possibility to interrupt activated molecular pathways. Far from being quiescent, the knee exists in state of chronic inflammation. In addition to reduced ROM knee fibrosis is a painful condition.\textsuperscript{29,34} CCL2 correlates positively with

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**Figure 3.** Elevated levels of vascular endothelial growth factor (VEGF), vascular cell adhesion molecule 1 (VCAM-1), and Intercellular adhesion molecule 1 (ICAM-1) in synovial fluid and tissue following primary total knee arthroplasty (TKA) in patients with joint fibrosis. No significant change in the levels of VEGF, VCAM-1, and ICAM-1 was found over time from primary TKA surgery in synovial fluid, fat pad, or synovial membrane tissue. The red line indicates the mean level in primary tissue for comparison. The black line represents the mean and black long dashed lines the 95% confidence interval. A linear regression model was used; $r^2$ and $p$ values are presented for each analysis [Color figure can be viewed at wileyonlinelibrary.com]
pain levels\textsuperscript{14}; blocking CCL2 signaling could reduce pain symptoms in these patients.

Although we report elevation of inflammatory mediators in revision patients many years from primary surgery, the question remains as to whether these mediators are important in pathogenesis of TKA failure. Longitudinal sampling is required to determine the contribution of these mediators to the pathogenesis of disease (e.g., fibrosis) that led to requirement of revision surgery. In addition, these mediators may be important in the 10–20% of patients that report unexplained knee pain and dissatisfaction following TKA.\textsuperscript{35} A longitudinal, prospective study is required to determine whether chronic inflammation is present in patients with unexplained pain not requiring revision surgery.

A number of biologically active markers involved in inflammation and matrix deposition were significantly elevated many years following primary TKA. Approved therapeutics are available that target these molecules, for example, CCL2, IL-8, and CCL3, and further studies are required to evaluate these compounds in patients with stiff, painful failed TKAs. In addition, prospective examination of the painful, stiff TKR is now required to determine (i) whether chronic inflammation is present in patients without a specific cause for their pain, and (ii) to identify molecular mediators to direct targeted immunomodulatory adjunctive therapy.

**AUTHORS’ CONTRIBUTION**

HLP, TB, CSG, ACP, NK, DJD, LB, and NK contributed to data acquisition, analysis, and interpretation. NK, LB, DJD, and DAM wrote the manuscript. All authors have read and approved the final submitted manuscript.

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REFERENCES


SUPPORTING INFORMATION

Additional supporting information may be found in the online version of this article.